

Metabolic, Nutritional, Iatrogenic, and Artifactual Sources of Urinary Organic Acids: A Comprehensive Table

ALAIN KUMPS, PIERRE DUEZ, and YVES MARDENS*

Background: The determination of organic acids and glycine conjugates in urine is key for the diagnosis and follow-up of several inborn errors of metabolism (IEM). However, clinical interpretations may still be hindered by ambiguity in the sources of some urinary organic acids and acylglycines as well as in the relationship between their excretion and IEM.

Approach: Relevant data have been compiled from major books and references on the topic and by exhaustive bibliographic searches through the Medline and Current Contents databases.

Content: A comprehensive table has been designed according to organic acids and conjugates. This table is intended to assist in the interpretation of organic acid profiles because, in addition to IEM, it also refers to other pathologic causes and to physiologic, nutritional, iatrogenic, and artifactual sources. Some preanalytical issues, including possible misinterpretations, are reviewed with regard to IEM.

© 2002 American Association for Clinical Chemistry

In the field of inborn errors of metabolism (IEM), “organic acids” are low-molecular weight (relative molecular weight less than ~300), water-soluble carboxylic acids that are intermediates or end products of amino acid, carbohydrate, lipid, or biogenic amine metabolism. Amino acids are excluded from this definition, whereas acylglycine conjugates and some decarboxylated derivatives are included because of their common clinical interest.

The analytical procedures for the determination of urinary organic acids usually include oximation, solvent extraction, and silylation followed by gas chromatogra-

phy with mass detection in scan mode data acquisition. Both the retention time and the mass spectrum allow the identification of the urinary metabolites, with quantification being performed on a specific fragment abundance (1–3). Analytical considerations can be found in the reports by Jellum (4), Chalmers and Lawson (1), Tuchman and Ulstrom (5), Niwa (6), Sweetman (2), and Duez et al. (3).

More than 250 organic acids and glycine conjugates are either typically present or may possibly be encountered in urine. More than 65 inherited metabolic abnormalities are known to yield a characteristic urinary organic acid pattern, essential for diagnosis and follow-up (1, 2, 5, 7, 8). The interpretation of urinary organic acid profiles can be difficult because of the variability of the compounds excreted. Moreover, there may still be a considerable degree of ambiguity in the origin and/or significance of a given compound. To arrive at a diagnosis, organic acid data can be correlated with, or confirmed by, other analyses, including plasma amino acid determination, plasma and cerebrospinal fluid lactate and pyruvate assays, whole blood acylcarnitine profiling, enzymatic activity determinations in blood cells or other cells, and genome analysis (7–11).

This report aims to compile information on the origins of the most frequently encountered urinary organic acids. In addition to IEM, our classification (Table 1) also refers to other pathologic conditions and physiologic, nutritional, iatrogenic, and artifactual causes (1, 2, 4–8, 10–13). This review is intended to assist in the interpretation of organic acid profiles and the identification of some preanalytical issues. Table 1, which is classified by organic compounds, is also proposed as a handy alternative that extends previously published compilations classified by inherited metabolic disorders (2, 5–7, 13).

Sampling Conditions

Urine collected over 24 h allows for variations in volume excretion during the day. The practicality of a 24-h collection is, however, such that a random specimen,

Laboratoire de Biochimie Médicale, Institut de Pharmacie, Université Libre de Bruxelles (ULB), Campus Plaine CP 205/3, Boulevard du Triomphe, B-1050 Brussels, Belgium.

*Author for correspondence. Fax 32-2-650-5324; e-mail biochmed@ulb.ac.be.

Received November 29, 2001; accepted January 25, 2002.

Table 1. Possible origins of abnormal excretion patterns of urinary organic acids.

Acid/Metabolite	Non-IEM (4, 12, 15, 16, 22)	IEM
Aromatic amino acid metabolism (23)		
2-Hydroxyphenylacetate	Uremia	PKU; BH4 ^a deficiency
4-Hydroxyphenylacetate (24, 25)	Bacterial gut metabolism and bacterial contamination (from tyrosine); short bowel syndrome; liver diseases	Tyrosinemia; PKU; hawkinsinuria
4-Hydroxyphenyllactate (24–27)	Bacterial gut metabolism; short bowel syndrome; liver diseases (e.g., secondary to PA, galactosemia, fructosemia); scurvy; lactic acidosis	Tyrosinemia; PKU; Zellweger; hawkinsinuria; lactic acidosis
4-Hydroxyphenylpyruvate	VPA; liver diseases (e.g., secondary to PA, galactosemia, fructosemia)	Tyrosinemia; hawkinsinuria
Homogentisate		Alcaptonuria
Mandelate (28)	Preservative in albumin solution for intravenous perfusion; methanamine mandelate; gastrointestinal malabsorption diseases	PKU
N-Acetyltyrosine	Some parenteral solutions	Tyrosinemia
Phenylacetate	Intestinal bacterial origin (from phenylalanine)	PKU; BH4 deficiency
Phenylacetylglutamine	Bacterial metabolism (from phenylacetate); hyperammonemia treated with phenylbutyrate or phenylacetate; uremia	PKU
Phenyllactate (29)	Bacterial gut metabolism (D-form); liver diseases	PKU; tyrosinemia (L-form); BH4 deficiency
Phenylpyruvate	Bacterial gut metabolism; liver diseases	PKU; BH4 deficiency
Succinylacetoacetate		Tyrosinemia type I
Succinylacetone		Tyrosinemia type I
Branched-chain amino acid metabolism		
2-Hydroxy-3-methylvalerate		MSUD; dihydrolipeoyl DH (E3) deficiency
2-Hydroxyisocaproate (23)	Short bowel syndrome (D-form)	MSUD; dihydrolipeoyl DH (E3) deficiency
2-Hydroxyisovalerate	Ketosis; lactic acidosis	MSUD; dihydrolipeoyl DH (E3) deficiency; MAD deficiency; lactic acidosis
2-Keto-3-methylvalerate	Lactic acidosis; ketosis	MSUD; dihydrolipeoyl DH (E3) deficiency; lactic acidosis
2-Ketoisocaproate	Lactic acidosis; ketosis	MSUD; dihydrolipeoyl DH (E3) deficiency; lactic acidosis
2-Ketoisovalerate	Lactic acidosis; ketosis	MSUD; dihydrolipeoyl DH (E3) deficiency; lactic acidosis
2-Methyl-acetoacetate (30)		Mitochondrial acetoacetyl-CoA-thiolase deficiency
2-Methylglutaconate		PA; MMA (?); β -ketothiolase deficiency
3-Hydroxy-2-ethylglutarate		PA
3-Hydroxy-2-ethylpropionate (31, 32)	Ketosis	3-Methylglutaconic aciduria (type II); methylmalonic semialdehyde DH deficiency ^c ; respiratory chain defects (complex I and II)
3-Hydroxy-2-methylbutyrate (30, 33–35)	Ketosis	Mitochondrial acetoacetyl-CoA-thiolase deficiency; 2-methyl-3-hydroxybutyryl-CoA DH deficiency; PA; Pearson syndrome
3-Hydroxy-3-methylglutarate	Ketosis	HMG-CoA lyase deficiency
3-Hydroxyisovalerate (30, 34)	Reye & Reye-like syndromes; VPA; ketosis	IVA; multicarboxylase deficiency; HMG-CoA lyase deficiency; 3-methylcrotonyl-CoA carboxylase deficiency; 3-methylglutaconyl-CoA hydratase deficiency; succinyl-CoA:3-oxoacid-CoA transferase deficiency; MAD deficiency
3-Hydroxypropionate (hydracrylate) (33, 34, 36)	Bacterial metabolism and contamination; short bowel syndrome; lactic acidosis	PA; MMA; multiple carboxylase deficiency; succinic semialdehyde DH deficiency; methylmalonic semialdehyde DH deficiency ^c ; lactic acidosis (with pyruvate carboxylase deficiency)
3-Keto-2-methylbutyrate (33)		PA; MMA (?); β -ketothiolase deficiency

Table 1. Continued.
Non-IEM (4, 12, 15, 16, 22)

Acid/Metabolite	Non-IEM (4, 12, 15, 16, 22)	IEM
3-Keto-2-methylvalerate (33)		PA; MMA (?); β -ketothiolase deficiency
3-Methylcrotonylglycine	Reye & Reye-like syndromes	3-Methylcrotonyl-CoA carboxylase deficiency; multiple carboxylase deficiency; HMG-CoA lyase deficiency
3-Methylglutaconate (37–42)	Uremia; acquired HMG-CoA lyase deficiency; other biochemical origin still unknown; pregnancy	3-Methylglutaconyl-CoA hydratase deficiency (methylglutaconic aciduria type I); HMG-CoA lyase deficiency; 3-methylglutaconic aciduria (other than type I); respiratory chain defects (e.g., Pearson syndrome or mitochondrial ATP synthase deficiency); Smith–Lemli–Opitz syndrome; carbamyl phosphate synthetase deficiency
3-Methylglutarate		As 3-methylglutaconate
4-Hydroxyisovalerate		IVA
Isovalerylglycine (34)	VPA	IVA; MAD deficiency; EMA aciduria (short-branched chain acyl-CoA DH deficiency; muscle COX deficiency ^a)
Methylcitrate (33, 34, 43)	Malnutrition	PA; MMA; multiple carboxylase deficiency
Methylmalonate (27, 34, 43–49)	B ₁₂ vitamin deficiency, pernicious anemia; bacterial gut metabolism; gastroenteritis in very young infants; short bowel syndrome; apnea; “benign” MMA; decreased GFR (in plasma); malnutrition	MMA; transcobalamin II deficiency; malonic aciduria
Propionylglycine (33, 34)		PA; MMA
Tiglylglycine (30, 31, 33–35, 50)	Reye & Reye-like syndromes; VPA	PA; 2-methyl-3-hydroxybutyryl-CoA DH deficiency; mitochondrial acetoacetyl-CoA-thiolase deficiency; multiple carboxylase deficiency; respiratory chain defects (e.g., complex I)
Fatty acid oxidation (16, 51–55)		
DCA (even, saturated): adipate, suberate, sebacate (17, 26, 43, 56–59)	Seriously ill states: infection, malnutrition, fever, seizures, liver diseases, pulmonary stenosis; MCT administration; ketosis; VPA or acetaminophen; lactic acidosis; hypoglycemia; Reye & Reye-like syndromes; Jamaican vomiting sickness	β -Oxidation defects (MAD, MCAD, SCAD, VLCAD, SCHAD, LCHAD/TFP); HMG-CoA lyase deficiency; systemic carnitine deficiency; succinic semialdehyde DH deficiency; CPT II deficiency; peroxisomal diseases; glycogen storage disorders I & II; lactic acidosis; fructose intolerance
Odd DCA (57, 58)	As even DCA; from plastic containers; uremia	Peroxisomal diseases
Unsaturated DCA (59)	Ketosis	VLCAD deficiency; CPT II deficiency
3-Hydroxy DCA (60)	MCT administration; fasting; ketosis; celiac disease	LCHAD/TFP deficiency; VLCAD deficiency
2-Hydroxysebacate (58)		Peroxisomal diseases
2-Methylbutylglycine	VPA	MAD deficiency; EMA aciduria (short-branched chain acyl-CoA DH deficiency; muscle COX deficiency ^a)
3-Hydroxyadipic (lactone)	See 3-hydroxy DCA	See 3-hydroxy DCA
3-Hydroxydo-/tetradecanedioate (61)	Hepatocellular disease; ketosis; acetaminophen intoxication	LCHAD/TFP deficiency
3-Hydroxysebacate (31, 62)	See 3-hydroxy DCA; progressive liver disease;	See 3-hydroxy DCA; MCAD deficiency; glycogen storage disorders I & II; secondary to respiratory chain defects
3-Hydroxysebacate	See 3-hydroxy DCA	See 3-hydroxy DCA
4-Octenedioate	Jamaican vomiting sickness; neonates on fasting	MCAD deficiency; MAD deficiency; VLCAD deficiency; LCHAD/TFP deficiency; nonketotic dicarboxyluria; systemic carnitine deficiency; peroxisomal diseases
5-Hydroxyhexanoate (59)	MCT administration; VPA; Reye & Reye-like syndromes; ketosis	MAD deficiency; MCAD deficiency; nonketotic dicarboxyluria
5-Hydroxysebacate		Peroxisomal diseases
7-Hydroxyoctanoate (59)	MCT administration; VPA	MCAD deficiency

Table 1. Continued.

Acid/Metabolite	Non-IEM (4, 12, 15, 16, 22)	IEM
Adipate	See DCA; food additive (Jello®); lithium; neonates on fasting	See DCA
Butyrylglycine	MCT administration; ketosis; Jamaican vomiting sickness	SCAD deficiency; MAD deficiency; EMA aciduria (short-branched chain acyl-CoA DH deficiency; muscle COX deficiency ⁶)
Decenedioate		MCAD deficiency; VLCAD deficiency; LCHAD/TFP deficiency
Do-/Tetradecanedioate (63)	Ketosis	VLCAD deficiency; LCHAD/TFP deficiency; MAD deficiency; CPT II deficiency
Ethylmalonate (11, 64–67)	Jamaican vomiting sickness; neonates on fasting; diet (?)	SCAD deficiency; MAD deficiency (severe form); MAD deficiency (mild form); acetyl-CoA carboxylase deficiency; EMA aciduria (short-branched chain acyl-CoA DH deficiency; muscle COX deficiency ⁶); respiratory chain defects
Hexanoylglycine (20, 59)	VPA; MCT administration; Jamaican vomiting sickness	MCAD deficiency; MAD deficiency; SCAD deficiency
Isobutyrylglycine		MAD deficiency; EMA aciduria (short-branched chain acyl-CoA DH deficiency, muscle COX deficiency ⁶)
Methylsuccinate		As EMA
Octanoate (59)	MCT administration	MCAD deficiency
Phenylpropionylglycine (20)	Bacterial gut metabolism and bacterial contamination	In MCAD deficiency (from phenylalanine bacterial metabolism or after load)
Suberylglycine (20, 59)	MCT administration; ketosis; Reye & Reye-like syndromes	MCAD deficiency; MAD deficiency
Tetradecanedioate		VLCAD deficiency; LCHAD/TFP deficiency; MAD deficiency
Krebs cycle/respiratory chain (68–71)		
2-Ketoglutarate (38, 72)	Bacterial contamination; lithium; uremia; increase with younger age	As malate; 2-ketoglutaric DH deficiency; GA I; 2-amino/2-ketoadipate acidemia; dihydrolipoyl DH (E3) deficiency; glycogen storage disorder I; 2-hydroxyglutaric aciduria (p-form); fumarase deficiency
Aconitate	High carbohydrate intake; parathyroid extract; saturnism; citrate intake; fruit juice added to urine;	Respiratory chain defects (e.g., complex I); Pearson syndrome
Citrate, isocitrate	hyperparathyroidism; increase with younger age	Dihydrolipoyl DH (E3) deficiency; fumarase deficiency; pyruvate carboxylase deficiency; Pearson syndrome
Fumarate	Lithium; renal tubular reabsorption defect (fumaric aciduria); increase with younger age	As malate; fumarase deficiency
Malate (73–75)	Lithium; uremia; increase with younger age	Respiratory chain defects; pyruvate carboxylase deficiency; PDH complex (E1, E3) deficiency; Pearson syndrome
Succinate (72, 76)	Bacterial (on storage); 2-ketoglutarate degradation; lithium; ketosis; tissue ischemia; increase with younger age	As malate; malonic aciduria; fumarase deficiency
Lactic acid, ketone bodies (30, 71, 77)		
2-Hydroxybutyrate	Ketosis; lactic acidosis	Lactic acidosis; GA I; respiratory chain defects
2-Hydroxyisobutyrate	Lactic acidosis	Lactic aciduria
3-Hydroxybutyrate (32, 49, 77–79)	Ketosis (e.g., vomiting, prolonged fasting, diabetic ketoacidosis); B ₁₂ vitamin deficiency; Reye & Reye-like syndromes; pulmonary infections; viral gastroenteritis; von Gierke disease; hyperthyroidism; pregnancy; heat stroke; ethanol; protein malnutrition; high-fat diet	Gluconeogenesis; PHD complex deficiency; respiratory chain defects; IVA; PA; MMA; multiple carboxylase deficiency; 3-methylcrotonyl-CoA carboxylase deficiency; glyceroluria; MSUD; GA I; MAD deficiency; β -ketothiolase deficiencies; 2-amino/2-ketoadipic acidemia; mitochondrial SCHAD; fatty acids oxidation deficiency (inappropriate ketosis)
Acetoacetate (78)	As 3-hydroxybutyrate; acetylsalicylate	As 3-hydroxybutyrate

Table 1. Continued.

Acid/Metabolite	Non-IEM (4, 12, 15, 16, 22)	IEM
Lactate and pyruvate (29, 78, 80–82)	Gut bacteria and bacterial contamination (D-lactate); short bowel syndrome (D-lactate); secondary lactic acidosis (e.g., apnea, septicemia, seizures, respiratory or cardiac insufficiency); diabetic ketoacidosis; Reye & Reye-like syndromes; increase with younger age; saccharose, fructose, lactose; drugs inducing hyperlactemia; dialysis bath; MCT administration	Primary lactic acidosis; PDH complex (E1, E2, E3) deficiency; oxidative phosphorylation and respiratory chain defects (e.g., MERRF, MELAS, Kearns-Sayre), Krebs acid cycle defects, gluconeogenesis defects (e.g., pyruvate carboxylase, fructose-1, 6-diphosphatase, glycogen storage I disorder); (short-branched chain acyl-CoA DH deficiency; muscle COX deficiency ^c); MAD deficiency (severe form); VLCAD deficiency; GA I; multiple carboxylase deficiency; some other organic aciduria (MMA, PA, IVA); citrullinemia, glycerol kinase deficiency; HMG-CoA lyase deficiency; EMA aciduria
Lysine, glycine, serine metabolism		
Glutaconate		GA I
Glycerate	Uremia; increase with younger age	D-Glyceric aciduria; hyperoxaluria type II (L-form); succinic semialdehyde DH deficiency
Glycolate (83, 84)	Ethylene glycol poisoning	Hyperoxaluria type I; succinic semialdehyde DH deficiency; isolated glycolic aciduria ^c
Glyoxylate		Hyperoxaluria type I
2-Hydroxyadipate (85)		2-Amino/2-Ketoadipic aciduria
3-Hydroxyglutarate (85)		GA I
2-Ketoadipate (85)		2-Amino/2-Ketoadipic aciduria
Glutarate (18, 19, 85–87)	2-Ketoglutarate degradation; bacterial gut metabolism; uremia; ethylene glycol poisoning; lithium	GA I; MAD deficiency (severe form); MAD deficiency (mild form); 2-amino/2-ketoadipic aciduria; malonic aciduria; other mitochondrial dysfunctions
Oxalate (83, 88–90)	Enteric malabsorption (regional enteritis or ileitis, celiac sprue disease, resection of ileum, Crohn disease); idiopathic stone disease; pyridoxine deficiency; increase with younger age; diet (e.g., beans, leafy vegetables, rhubarb, spinach, tomatoes, strawberries, tea, chocolate); infant formula; ascorbic acid; xylitol; ethylene glycol; methoxyflurane	Hyperoxaluria type I and II; hyperoxaluria without known enzyme deficit
Other acids and metabolites		
2-Hydroxyglutarate (82, 85)	Bacterial contamination (D-form); lithium; uremia; increase with younger age; 2-ketoglutarate degradation	2-Hydroxyglutaric aciduria (L- and D-forms); MAD deficiency, severe (D-form); MAD deficiency, mild (D-form); 2-amino/2-ketoadipic aciduria; malonic aciduria
3,4-Dihydroxybutyrate (2-deoxytetronate)	Diet	Succinic semialdehyde DH deficiency
3-Hydroxyisobutyrate (32)	Ketosis	3-Hydroxyisobutyric DH deficiency and/or methylmalonic semialdehyde DH deficiency ^c
4,5-Dihydroxyhexanoate	Ketosis (?)	Succinic semialdehyde DH deficiency
4-Hydroxybutyrate	Bacterial gut metabolism (?)	Succinic semialdehyde DH deficiency
4-Hydroxycyclohexylacetate (91)	Contamination (suppository, emollients); uremia	Hawkinsinuria
Glycerol		Glycerol kinase deficiency; fructose-1,6-phosphatase deficiency
Malonate (92)		Malonyl-CoA-decarboxylase deficiency; malonic aciduria with normal malonyl-CoA-decarboxylase activity
Mevalonate and/or its lactone		Mevalonate kinase deficiency
N-Acetylaspartate		Canavan disease
Orotate (93–95)	Allopurinol treatment; azauridine; high cell turnover (tissue breakdown, menstruation); folate malabsorption	Argininemia; orotic aciduria; citrullinemia; OCT deficiency; hyperornithinemia-hyperammonemia-homocitrullinuria syndrome; lysinuric protein intolerance; purine nucleoside phosphorylase deficiency; Lesh–Nyhan disease

Table 1. Continued.

Acid/Metabolite	Non-IEM (4, 12, 15, 16, 22)	IEM
Pyroglutamate (L- or D-5-oxoproline) (82, 91, 96–102)	From glutamine of hydrolyzed proteins (infant formula); acetaminophen; vigabatrin; fludoxacin, netilmicin (?); glutamine degradation (in hyperammonemia, urea cycle defects); vegetarian or low-protein diets, undernutrition; iron oxoprolinate; Steven–Johnson syndrome; burns; premature newborns; transitory (?); glycine deficiency; increase with younger age; renal insufficiency; pregnancy (increased metabolic demand for glycine)	Glutathione synthetase deficiency; 5-oxoprolinase deficiency; nephropathic cystinosis; hawkinsinuria; homocystinuria; OCT deficiency; PA
Thymine	Caffeine (?)	Dihydropyrimidine DH deficiency
Uracil	Caffeine (?)	Dihydropyrimidine DH deficiency; OCT deficiency; citrullinemia
Vanillactate (103)	Catecholamine-containing foodstuff (e.g., bananas); L-dopa decarboxylase inhibitors; neuroblastoma	L-Amino acid decarboxylase deficiency
Nutritional, exogenous, or artifactual compounds (6)		
2,5-Furane dicarboxylate and 5-hydroxymethyl-2-furanoate	Heated furanoic sugars (chocolates, fruit juice, intravenous perfusion)	
2-Furoylglycine	Chocolate; heated fruit juice or parenteral solution; uremia	
3-(3-Hydroxyphenyl)hydracrylate	From nutrition	
4-Hydroxycyclohexane-1-carboxylate (104)	Diet; bacterial gut metabolism (from tyrosine)	
4-Hydroxyhippurate	Bacterial gut metabolism	
Benzoate (61, 105)	Bacterial metabolism (gut, urinary tract) from hippurate or from aromatic amino acids; benzoate treatment; food additive; ethylene glycol poisoning; toluene; hyperammonemia	
Hippurate (106)	As benzoate; uremia	
Maleate	Fluvoxamine maleate	
Palmitate	Soap; Jamaican vomiting sickness	
<i>p</i> -Cresol	Bacterial metabolism from tyrosine; toluene; uremia	
Phenol	Bacterial metabolism from tyrosine; exposure to benzene or phenol; malabsorption; uremia	
Pivalate	Pivampicillin or pivmecillinam	
Tartarate	Food additive; uremia	
<p>^a PKU, phenylketonuria; BH4, tetrahydrobiopterin; PA, propionic acidemia; VPA, valproate; MSUD, maple syrup urine disease; DH, dehydrogenase; MMA, methylmalonic acidemia; HMG, 3-hydroxy-3-methylglutarate; IVA, isovaleric acidemia; MAD, multiple acyl-CoA dehydrogenase; EMA, ethylmalonate; COX, cytochrome c oxidase; GFR, glomerular filtration rate; DCA, dicarboxylic acid; MCT, medium-chain triglyceride; MCAD, medium-chain acyl-CoA dehydrogenase; SCAD, short-chain acyl-CoA dehydrogenase; VLCAD, very long-chain acyl-CoA dehydrogenase; SCHAD, short-chain 3-hydroxyacyl-CoA dehydrogenase; LCHAD/TFP, long-chain 3-hydroxyacyl-CoA dehydrogenase/trifunctional protein; CPT, carnitine palmitoyltransferase; GA I, glutaric aciduria type I; PDH, pyruvate dehydrogenase; OCT, ornithine carbamoyltransferase.</p> <p>^b (?) indicates that this information remains to be confirmed.</p> <p>^c Enzymatic confirmation not yet established.</p>		

preferably the first morning voiding, is an acceptable alternative. This specimen usually consists of at least 2 mL and is stored until analysis at below -18°C without the use of any preservative.

Intraindividual variations will occur with respect to the time of sampling, the patient's clinical status, eventual diet management, and whether the sample is collected when the patient is fasted or fed. Sampling during fasting or metabolic decompensation is often considered to be most valuable because, in most cases, metabolites of interest are then excreted selectively or at a higher concentration. On the other hand, metabolic decompensation, such as lactic acidosis, ketosis, or liver failure, gives rise to an abnormal excretion of organic acids (α -keto branched, dicarboxylic, or aromatic acids, respectively) that are otherwise involved in particular IEM; this sometimes renders interpretation even more difficult.

Poor preservation of samples will lead to nonenzymatic conversion of all keto acids to the respective hydroxyacid; for example, acetoacetate is converted to 3-hydroxybutyrate, and 2-ketoglutarate is converted to 2-hydroxyglutarate.

Abnormal Excretion Patterns Not Attributable to IEM

An increase in excretion may be nonspecific because some metabolites are reported to be abnormally excreted in conditions not attributable to IEM (drug therapy, diet, non-IEM diseases, or physiologic conditions), as indicated in Table 1.

Two frequent abnormal excretions not necessarily related to IEM are lactic aciduria and ketonuria. Whatever its origin, lactic aciduria is generally accompanied by other compounds; the greater the lactate excretion, the more likely the extent of the excretion of pyruvate, *p*-hydroxyphenyllactate, 2-hydroxyisovalerate, 2-hydroxybutyrate, and to a lesser extent, branched-chain 2-ketoacids. The abnormal excretion of these branched compounds implies the need for differentiation from dihydrolipoyl dehydrogenase deficiency.

Ketonuria (3-hydroxybutyrate and acetoacetate) is often accompanied by 3-hydroxyisobutyrate, 3-hydroxyisovalerate, 2-hydroxybutyrate, and dicarboxylic acids, particularly their 3-hydroxy derivatives with chain lengths up to C14. In this latter case, the pattern could mimic a long-chain 3-hydroxyacyl-CoA dehydrogenase or a trifunctional protein deficiency profile, except for the very high excretion of ketone bodies [in fatty acid oxidation defects, ketone bodies may appear increased in urine during fasting, but the ketosis remains at an inappropriately low level and the ratio of urinary adipate to 3-hydroxybutyrate is >0.5 (14)].

Another common misinterpretation may arise from bacterial metabolism. Of possible endogenous origin (e.g., intestinal infection) is the abnormal excretion of D-lactate (not chromatographically separated from L-lactate), methylmalonate, *p*-hydroxyphenylacetate, *p*-hydroxyphenyllactate, phenylacetylglutamine, phenylpropionylglycine,

glutarate, benzoate, and hippurate. Of possible exogenous origin (bacterial growth in urine) are D-lactate, 2-ketoglutarate, D-2-hydroxyglutarate, succinate, 3-hydroxypropionate, and phenol derivatives (phenol, *p*-cresol, hippurate) (15).

The drug valproic acid may lead to increased excretion of 3-hydroxyisovalerate, 5-hydroxyhexanoate, 7-hydroxyoctanoate, *p*-hydroxyphenylpyruvate, dicarboxylic acids, and to a lesser extent, hexanoylglycine, tiglylglycine, and isovalerylglycine. The metabolites of this anticonvulsant drug are an important clue to the analyst, however.

The administration of medium-chain triglycerides may yield a pattern resembling fatty acid β -oxidation defects, with increased saturated even-numbered dicarboxylic acids, mainly sebacate, as well as increased 5-hydroxyhexanoate and 7-hydroxyoctanoate, and the presence of octanoate but the absence or low excretion of glycine derivatives (16, 17).

Misleading Normal or Near-Normal Excretion

The excretion of organic acids in pathologic conditions may be characterized by large variability and thus casts doubt on the clinical sensitivity of the results. Interindividual variations are also possible because, for some diseases, urinary biochemical features may depend on what have been called "excretory" and "non-excretory" patients. Indeed, compounds typically excreted in large amounts may also appear at only slightly increased or even normal concentrations in some IEM. This is particularly true when a patient is clinically well (not in a state of metabolic decompensation) or under suitable dietary control. Among these inborn errors are glutaric aciduria type I (18, 19) (glutarate concentrations may be within reference values, whereas 3-hydroxyglutarate is present); medium-chain acyl-CoA dehydrogenase deficiency (adipate, suberate, and sebacate concentrations may be within reference values, but the presence of suberylglycine and hexanoylglycine will reveal the disorder) (20); multiple acyl-CoA dehydrogenase deficiency, particularly in its mild forms (metabolites suggesting such a disease, including ethylmalonate and glutarate, are quite variable); and 2-ketoglutarate dehydrogenase deficiency (2-ketoglutarate excretion ranges from within reference values to 10 times higher than the upper limit of the reference interval).

Respiratory chain defects give an unpredictable organic acid pattern, but nearly always with marked lactic aciduria; Krebs cycle acids, ethylmalonate, 3-methylglutaconate, and 3-methylglutarate may also be excreted in varying quantities. Urinary orotate may be high but possibly borderline in citrullinemia, ornithine carbamoyltransferase deficiency, lysinuric protein intolerance, and the hyperornithinemia-hyperammonemia-homocitrullinuria syndrome, all disorders for which the biochemical diagnosis, however, is based on plasma ammonium and plasma and urinary amino acid profiles.

Interpretation and Misinterpretations (7, 8, 10, 11)

The relevance of the abnormal excretion of some characteristic metabolites in the diagnosis of IEM has to be emphasized. For example, the presence of succinylacetone and succinylacetoacetate is pathognomonic of tyrosinemia type I (fumarylacetoacetate hydrolase deficiency). Other compounds may also be quite specific, including 3-hydroxyglutarate for glutaric aciduria type I, mevalonic acid for mevalonic aciduria, *N*-acetylaspartate for Canavan disease, 4-hydroxycyclohexylacetate for hawkinsinuria, and 2-ketoadipate and 2-hydroxyadipate for 2-amino/2-ketoadipate aciduria.

Cooperation between clinical chemists and clinicians is essential for the interpretation of the results. On the one hand, information on diet, drug intake, and clinical symptoms and signs may often be required by the clinical chemist to refine his or her interpretation. The clinical chemist can inform the clinician of pitfalls, the possible origins of abnormal results, and further analyses that can be performed (21). On the other hand, a final diagnosis can be established only in terms of the patient's history and clinical picture, in addition to results from biochemical and medical examinations.

CONCLUSION

As a practical consequence of possible misinterpretations, urinary organic acid patterns must be interpreted in the context of the complete clinical picture. In this context, both an abnormal organic acid pattern in the urine from an asymptomatic individual and a normal profile from a patient suspected of IEM must be considered as indications for repeated sampling: in the former circumstance, more information on possible drug therapy, diet, non-IEM pathology, and physiologic conditions is mandatory, whereas in the latter case, a period of illness would be preferred for resampling.

References

- Chalmers RA, Lawson AM. Organic acids in man. London; New York: Chapman and Hall, 1982:524pp.
- Sweetman L. Organic acid analysis. In: Hommes FA, ed. Techniques in diagnostic human biochemical genetics. A laboratory manual. New York: Wiley-Liss, 1991:143–76.
- Duez P, Kumps A, Mardens Y. GC-MS profiling of urinary organic acids evaluated as a quantitative method. *Clin Chem* 1996;42:1609–15.
- Jellum E. Profiling of human body fluids in healthy and diseased states using gas chromatography and mass spectrometry, with special reference to organic acids. *J Chromatogr* 1977;143:427–62.
- Tuchman M, Ulstrom RA. Urinary organic acids in health and disease. *Adv Pediatr* 1985;32:469–506.
- Niwa T. Metabolic profiling with gas chromatography-mass spectrometry and its application to clinical medicine. *J Chromatogr* 1986;379:313–45.
- Blau N, Duran M, Blaskovics ME. Physician's guide to the laboratory diagnosis of metabolic diseases. London: Chapman & Hall, 1996:508pp.
- Scriver CR, Beaudet AL, Sly WS, Valle D. The metabolic and molecular bases of inherited diseases, 8th ed. New York: McGraw-Hill, 2001:6484pp.
- Clarke JTR. A clinical guide to inherited metabolic diseases. Cambridge: Cambridge University Press, 1996:280pp.
- Fernandes J, Saudubray J-M, Van den Berghe G. Inborn metabolic diseases: diagnosis and treatment, 3rd revised ed. Berlin: Springer-Verlag, 2000:468pp.
- Nyhan WL, Ozand PT. Atlas of metabolic diseases. London: Chapman & Hall Medical, 1998:680pp.
- Young DS. Effects of drugs on clinical laboratory tests, 5th ed. Washington: AACC Press, 2000:2200pp.
- Friedman RB, Young DS. Effects of disease on clinical laboratory tests, 3rd ed. Washington: AACC Press, 1997:1068pp.
- Treacy E, Pitt J, Egginton M, Hawkins R. Dicarboxylic aciduria, significance and prognostic indications. *Eur J Pediatr* 1994;153:918.
- Hansen S, Perry TL, Lesk D, Gibson L. Urinary bacteria: potential source of some organic acidurias. *Clin Chim Acta* 1972;39:71–4.
- Bohles H, Akcetin Z, Lehnert W. The influence of intravenous medium- and long-chain triglycerides and carnitine on the excretion of dicarboxylic acids. *J Parenter Enteral Nutr* 1987;11:46–8.
- Mortensen PB, Gregersen N. Medium-chain triglyceride medication as a pitfall in the diagnosis of non-ketotic C6–C10-dicarboxylic acidurias. *Clin Chim Acta* 1980;103:33–7.
- Campistol J, Ribes A, Alvarez L, Christensen E, Millington DS. Glutaric aciduria type I: unusual biochemical presentation. *J Pediatr* 1992;121:83–6.
- Baric I, Wagner L, Feyh P, Liesert M, Buckel W, Hoffmann GF. Sensitivity and specificity of free and total glutaric acid and 3-hydroxyglutaric acid measurements by stable-isotope dilution assays for the diagnosis of glutaric aciduria type I. *J Inher Metab Dis* 1999;22:867–81.
- Rinaldo P, O'Shea JJ, Coates PM, Hale DE, Stanley CA, Tanaka K. Medium-chain acyl-CoA dehydrogenase deficiency. Diagnosis by stable-isotope dilution measurement of urinary *n*-hexanoylglycine and 3-phenylpropionylglycine. *N Engl J Med* 1988;319:1308–13.
- Blom W, Huijman JG, van den Berg GB. A clinical biochemist's view of the investigation of suspected inherited metabolic disease. *J Inher Metab Dis* 1989;12:64–88.
- Heindl A, Dietel P, Spittler G. Distinction between urinary acids originating from nutrition and those produced in the human body. *J Chromatogr* 1986;377:3–14.
- Spaapen LJ, Ketting D, Wadman SK, Bruinvis L, Duran M. Urinary *D*-4-hydroxyphenyllactate, *D*-phenyllactate and *D*-2-hydroxyisocaproate, abnormalities of bacterial origin. *J Inher Metab Dis* 1987;10:383–90.
- van der Heiden C, Wauters EA, Duran M, Wadman SK, Ketting D. Gas chromatographic analysis of urinary tyrosine and phenylalanine metabolites in patients with gastrointestinal disorders. *Clin Chim Acta* 1971;34:289–96.
- Chalmers RA, Valman HB, Liberman MM. Measurement of 4-hydroxyphenylacetic aciduria as a screening test for small-bowel disease. *Clin Chem* 1979;25:1791–4.
- Mayatepek E, Seppel CK, Hoffmann GF. Increased urinary excretion of dicarboxylic acids and 4-hydroxyphenyllactic acid in patients with Zellweger syndrome. *Eur J Pediatr* 1995;154:755–6.
- Tuchman M, McCann MT, Thompson MM, Tsai MY, Giguere R, Lemieux B. Screening urine of 3-week-old newborns: transient methylmalonic and hydroxyphenyllactic aciduria. *Biochem Med Metab Biol* 1992;48:64–8.
- Hill A, Hoag GN, Zaleski WA. The investigation of aromatic acids in phenylketonuria, alkaptonuria and tyrosinosis using gas-liquid chromatography. *Clin Chim Acta* 1972;37:455–62.
- Heil M, Podebrad F, Beck T, Mosandl A, Sewell AC, Bohles H.

- Enantioselective multidimensional gas chromatography-mass spectrometry in the analysis of urinary organic acids. *J Chromatogr B Biomed Sci Appl* 1998;714:119–26.
30. Mitchell GA, Fukao T. Inborn errors of ketone body metabolism. In: Scriver CR, Beaudet AL, Sly WS, Valle D, eds. *The metabolic and molecular bases of inherited diseases*, 8th ed. New York: McGraw-Hill, 2001:2327–56.
 31. Bennett MJ, Weinberger MJ, Sherwood WG, Burlina AB. Secondary 3-hydroxydicarboxylic aciduria mimicking long-chain 3-hydroxyacyl-CoA dehydrogenase deficiency. *J Inherit Metab Dis* 1994;17:283–6.
 32. Gibson KM, Lee CF, Bennett MJ, Holmes B, Nyhan WL. Combined malonic, methylmalonic and ethylmalonic acid semialdehyde dehydrogenase deficiencies: an inborn error of β -alanine, L-valine and L-alloisoleucine metabolism? *J Inherit Metab Dis* 1993;16:563–7.
 33. Lehnert W, Spertl W, Suormala T, Baumgartner ER. Propionic acidemia: clinical, biochemical and therapeutic aspects. Experience in 30 patients. *Eur J Pediatr* 1994;153:S68–80.
 34. Ogier de Baulny H, Saudubray J-M. Branched-chain organic acidurias. In: Fernandes J, Saudubray J-M, Van den Berghe G, eds. *Inborn metabolic diseases. Diagnosis and treatment*, 3rd ed. Berlin: Springer-Verlag, 2000:195–212.
 35. Zschocke J, Ruitter JP, Brand J, Hoffmann GF, Wanders RJ, Mayatepek E. Progressive infantile neurodegeneration caused by 2-methyl-3-hydroxybutyryl-CoA dehydrogenase deficiency: a novel inborn error of branched-chain fatty acid and isoleucine metabolism. *Pediatr Res* 2000;48:852–5.
 36. Pollitt RJ, Fowler B, Sardharwalla IB, Edwards MA, Gray RG. Increased excretion of propan-1,3-diol and 3-hydroxypropionic acid apparently caused by abnormal bacterial metabolism in the gut. *Clin Chim Acta* 1987;169:151–7.
 37. Gibson KM, Sherwood WG, Hoffman GF, Stumpf DA, Dianzani I, Schutgens RB, et al. Phenotypic heterogeneity in the syndromes of 3-methylglutaconic aciduria. *J Pediatr* 1991;118:885–90.
 38. Guffon N, Lopez-Mediavilla C, Dumoulin R, Mousson B, Godinot C, Carrier H, et al. 2-Ketoglutarate dehydrogenase deficiency, a rare cause of primary hyperlactataemia: report of a new case. *J Inherit Metab Dis* 1993;16:821–30.
 39. Kelley RI, Kratz L. 3-Methylglutaconic acidemia in Smith-Lemli-Opitz syndrome. *Pediatr Res* 1995;37:671–4.
 40. Gibson KM, Bennett MJ, Mize CE, Jakobs C, Rotig A, Munnich A, et al. 3-Methylglutaconic aciduria associated with Pearson syndrome and respiratory chain defects. *J Pediatr* 1992;121:940–2.
 41. Mills GA, Hill MA, Buchanan R, Corina DL, Walker V. 3-Hydroxy-3-methylglutaric aciduria: a possible pitfall in diagnosis. *Clin Chim Acta* 1991;204:131–6.
 42. Walsh R, Conway H, Roche G, Mayne PD. What is the origin of 3-methylglutaconic acid? *J Inherit Metab Dis* 1999;22:251–5.
 43. Teran-Garcia M, Ibarra I, Velazquez A. Urinary organic acids in infant malnutrition. *Pediatr Res* 1998;44:386–91.
 44. Rasmussen K, Vyberg B, Pedersen KO, Brochner-Mortensen J. Methylmalonic acid in renal insufficiency: evidence of accumulation and implications for diagnosis of cobalamin deficiency. *Clin Chem* 1990;36:1523–4.
 45. Stabler SP, Marcell PD, Podell ER, Allen RH, Lindenbaum J. Assay of methylmalonic acid in the serum of patients with cobalamin deficiency using capillary gas chromatography-mass spectrometry. *J Clin Invest* 1986;77:1606–12.
 46. Treacy E, Clow C, Mamer OA, Scriver CR. Methylmalonic acidemia with a severe chemical but benign clinical phenotype. *J Pediatr* 1993;122:428–9.
 47. Sewell AC, Herwig J, Bohles H. A case of familial 'benign' methylmalonic aciduria? *J Inherit Metab Dis* 1996;19:696–7.
 48. Artuch R, Calvo M, Ribes A, Camarasa F, Vilaseca MA. Increased urine methylmalonic acid excretion in infants with apnoeas. *J Inherit Metab Dis* 1998;21:86–7.
 49. Nowaczyk MJM, Lehotay DC, Platt BA, Clarke JTR. Urinary organic acid profiles in infants during acute illness. In: Society for Inherited Metabolic Disorders, 7th International Congress of Inborn Errors of Metabolism, Vienna, Austria, 1997:102.
 50. Bennett MJ, Sherwood WG, Gibson KM, Burlina AB. Secondary inhibition of multiple NAD-requiring dehydrogenases in respiratory chain complex I deficiency: possible metabolic markers for the primary defect. *J Inherit Metab Dis* 1993;16:560–2.
 51. Duran M, Wadman SK. Chemical diagnosis of inherited defects of fatty acid metabolism and ketogenesis. *Enzyme* 1987;38:115–23.
 52. Hale DE, Bennett MJ. Fatty acid oxidation disorders: a new class of metabolic diseases. *J Pediatr* 1992;121:1–11.
 53. Rhead WJ. Inborn errors of fatty acid oxidation in man. *Clin Biochem* 1991;24:319–29.
 54. Whyte RK, Whelan D, Hill R, McClorry S. Excretion of dicarboxylic and omega-1 hydroxy fatty acids by low birth weight infants fed with medium-chain triglycerides. *Pediatr Res* 1986;20:122–5.
 55. Stanley CA. Disorders of fatty acid oxidation. In: Fernandes J, Saudubray J-M, Van den Berghe G, eds. *Inborn metabolic diseases. Diagnosis and treatment*, 3rd ed. Berlin: Springer-Verlag, 2000:139–50.
 56. Bergoffen J, Kaplan P, Hale DE, Bennett MJ, Berry GT. Marked elevation of urinary 3-hydroxydecanedioic acid in a malnourished infant with glycogen storage disease, mimicking long-chain L-3-hydroxyacyl-CoA dehydrogenase deficiency. *J Inherit Metab Dis* 1993;16:851–6.
 57. Rocchiccioli F, Aubourg P, Bougneres PF. Medium- and long-chain dicarboxylic aciduria in patients with Zellweger syndrome and neonatal adrenoleukodystrophy. *Pediatr Res* 1986;20:62–6.
 58. Rizzo C, Bertucci P, Federici G, Wanders RJA, Sabetta G, Dionisi-Vici C. Urinary organic acids in peroxisomal disorders. *J Inherit Metab Dis* 2000;23(Suppl 1):241.
 59. Yamaguchi S, Shimizu N, Orii T, Fukao T, Suzuki Y, Maeda K, et al. Prenatal diagnosis and neonatal monitoring of a fetus with glutaric aciduria type II due to electron transfer flavoprotein (β -subunit) deficiency. *Pediatr Res* 1991;30:439–43.
 60. Costa CG, Verhoeven NM, Kneepkens CM, Douwes AC, Wanders RJ, et al. Organic acid profiles resembling a β -oxidation defect in two patients with coeliac disease. *J Inherit Metab Dis* 1996;19:177–80.
 61. Nowaczyk MJ, Whelan D, Hill RE, Clarke JT, Pollitt RJ. Long-chain hydroxydicarboxylic aciduria, carnitine depletion and acetaminophen exposure. *J Inherit Metab Dis* 2000;23:188–9.
 62. Greter J, Lindstedt S, Seeman H, Steen G. 3-Hydroxydecanedioic acid and related homologues: urinary metabolites in ketoacidosis. *Clin Chem* 1980;26:261–5.
 63. Jackson S, Bartlett K, Land J, Moxon ER, Pollitt RJ, Leonard JV, et al. Long-chain 3-hydroxyacyl-CoA dehydrogenase deficiency. *Pediatr Res* 1991;29:406–11.
 64. Ozand PT, Rashed M, Millington DS, Sakati N, Hazzaa S, Rahbeeni Z, et al. Ethylmalonic aciduria: an organic acidemia with CNS involvement and vasculopathy. *Brain Dev* 1994;16(Suppl):12–22.
 65. Christensen E, Brandt NJ, Schmalbruch H, Kamieniecka Z, Hertz B, Ruitenbeek W. Muscle cytochrome c oxidase deficiency accompanied by a urinary organic acid pattern mimicking multiple acyl-CoA dehydrogenase deficiency. *J Inherit Metab Dis* 1993;16:553–6.
 66. Lehnert W, Ruitenbeek W. Ethylmalonic aciduria associated with progressive neurological disease and partial cytochrome c oxidase deficiency. *J Inherit Metab Dis* 1993;16:557–9.
 67. Burlina AB, Dionisi-Vici C, Bennett MJ, Gibson KM, Servidei S,

- Bertini E, et al. A new syndrome with ethylmalonic aciduria and normal fatty acid oxidation in fibroblasts. *J Pediatr* 1994;124:79–86.
68. Ribes A, Riudor E, Valcarel R, Salva A, Castello F, Murillo S, et al. Pearson syndrome: altered tricarboxylic acid and urea-cycle metabolites, adrenal insufficiency and corneal opacities. *J Inher Metab Dis* 1993;16:537–40.
69. Munnich A, Rotig A, Chretien D, Saudubray J-M, Cormier V, Rustin P. Clinical presentations and laboratory investigations in respiratory chain deficiency. *Eur J Pediatr* 1996;155:262–74.
70. Bauer MF, Gempel K, Hofmann S, Jaksch M, Philbrook C, Gerbitz KD. Mitochondrial disorders. A diagnostic challenge in clinical chemistry. *Clin Chem Lab Med* 1999;37:855–76.
71. Kerr DS, Wexler ID, Zinn AB. Disorders of pyruvate metabolism and the tricarboxylic acid cycle. In: Fernandes J, Saudubray J-M, Van den Berghe G, eds. *Inborn metabolic diseases. Diagnosis and treatment*, 3rd ed. Berlin: Springer-Verlag, 2000:126–38.
72. Rustin P, Bourgeron T, Parfait B, Chretien D, Munnich A, Rotig A. Inborn errors of the Krebs cycle: a group of unusual mitochondrial diseases in human. *Biochim Biophys Acta* 1997;1361:185–97.
73. Ederly P, Gerard B, Chretien D, Rotig A, Cerrone R, Rabier D, et al. Liver cytochrome c oxidase deficiency in a case of neonatal-onset hepatic failure. *Eur J Pediatr* 1994;153:190–4.
74. Das AM, Schweitzer-Krantz S, Byrd DJ, Brodehl J. Absence of cytochrome c oxidase activity in a boy with dysfunction of renal tubules, brain and muscle. *Eur J Pediatr* 1994;153:267–70.
75. Caruso U, Adami A, Bertini E, Burlina AB, Carnevale F, Cerone R, et al. Respiratory-chain and pyruvate metabolism defects: Italian collaborative survey on 72 patients. *J Inher Metab Dis* 1996;19:143–8.
76. Liebich HM, Pickert A, Stierle U, Woll J. Gas chromatography-mass spectrometry of saturated and unsaturated dicarboxylic acids in urine. *J Chromatogr* 1980;199:181–9.
77. Liebich HM. Gas chromatographic profiling of ketone bodies and organic acids in diabetes. *J Chromatogr* 1986;379:347–66.
78. Vassault A, Bonnefont JP, Specola N, Saudubray J-M. Lactate, pyruvate, and ketone bodies. In: Hommes FA, ed. *Techniques in diagnostic human biochemical genetics. A laboratory manual*. New York: Wiley-Liss, 1991:285–308.
79. Bennett MJ, Weinberger MJ, Kabori JA, Rinaldo P, Burlina AB. Mitochondrial short-chain L-3-hydroxyacyl-coenzyme A dehydrogenase deficiency: a new defect of fatty acid oxidation. *Pediatr Res* 1996;39:185–8.
80. Madias NE. Lactic acidosis. *Kidney Int* 1986;29:752–74.
81. Bongaerts G, Tolboom J, Naber T, Bakkeren J, Severijnen R, Willems H. D-Lactic acidemia and aciduria in pediatric and adult patients with short bowel syndrome. *Clin Chem* 1995;41:107–10.
82. Sewell AC, Heil M, Podebrad F, Mosandl A. Chiral compounds in metabolism: a look in the molecular mirror. *Eur J Pediatr* 1998;157:185–91.
83. Latta K, Brodehl J. Primary hyperoxaluria type I. *Eur J Pediatr* 1990;149:518–22.
84. Craigen WJ. Persistent glycolic aciduria in a healthy child with normal alanine-glyoxylate aminotransferase activity. *J Inher Metab Dis* 1996;19:793–4.
85. Hoffmann GF. Disorders of lysine catabolism and related cerebral organic-acid disorders. In: Fernandes J, Saudubray J-M, Van den Berghe G, eds. *Inborn metabolic diseases. Diagnosis and treatment*, 3rd ed. Berlin: Springer-Verlag, 2000:241–53.
86. Haworth JC, Booth FA, Chudley AE, deGroot GW, Dilling LA, Goodman SI, et al. Phenotypic variability in glutaric aciduria type I: report of fourteen cases in five Canadian Indian kindreds. *J Pediatr* 1991;118:52–8.
87. Wendel U, Bakkeren J, de Jong J, Bongaerts G. Glutaric aciduria mediated by gut bacteria. *J Inher Metab Dis* 1995;18:358–9.
88. Chalmers RA, Purkiss P. Oxalic acid in plasma and urine. In: Hommes FA, ed. *Techniques in diagnostic human biochemical genetics. A laboratory manual*. New York: Wiley-Liss, 1991:359–76.
89. Searcy RL. Urolithiasis and the oxalate answer. *Int Clin Prod Rev* 1986;18–22.
90. Van Acker KJ, Eyskens FJ, Espeel MF, Wanders RJ, Dekker C, Kerckaert IO, et al. Hyperoxaluria with hyperglycoluria not due to alanine:glyoxylate aminotransferase defect: a novel type of primary hyperoxaluria. *Kidney Int* 1996;50:1747–52.
91. Hocart CH, Halpern B, Hick LA, Wong CO. Hawkinsinuria—identification of quinolacetic acid and pyroglutamic acid during an acidotic phase. *J Chromatogr* 1983;275:237–43.
92. Gregg AR, Warman AW, Thorburn DR, O'Brien WE. Combined malonic and methylmalonic aciduria with normal malonyl-coenzyme A decarboxylase activity: a case supporting multiple aetiologies. *J Inher Metab Dis* 1998;21:382–90.
93. Burlina AB, Ferrari V, Dionisi-Vici C, Bordugo A, Zacchello F, Tuchman M. Allopurinol challenge test in children. *J Inher Metab Dis* 1992;15:707–12.
94. Corbeel L, Van den Berghe G, Jaeken J, Van Tornout J, Eeckels R. Congenital folate malabsorption. *Eur J Pediatr* 1985;143:284–90.
95. Brusilow SW. Determination of urine orotate and orotidine and plasma ammonium. In: Hommes FA, ed. *Techniques in diagnostic human biochemical genetics. A laboratory manual*. New York: Wiley-Liss, 1991:345–57.
96. Oberholzer VG, Wood CB, Palmer T, Harrison BM. Increased pyroglutamic acid levels in patients on artificial diets. *Clin Chim Acta* 1975;62:299–304.
97. Nakanishi T, Shimizu A, Saiki K, Fujiwara F, Funahashi S, Hayashi A. Quantitative analysis of urinary pyroglutamic acid in patients with hyperammonemia. *Clin Chim Acta* 1991;197:249–55.
98. Grimble G. Glutamine, glutamate and pyroglutamate—facts and fantasies. *Clin Nutr* 1993;12:66–9.
99. Croal BL, Glen AC, Kelly CJ, Logan RW. Transient 5-oxoprolinuria (pyroglutamic aciduria) with systemic acidosis in an adult receiving antibiotic therapy. *Clin Chem* 1998;44:336–40.
100. Jackson AA, Badaloo AV, Forrester T, Hibbert JM, Persaud C. Urinary excretion of 5-oxoproline (pyroglutamic aciduria) as an index of glycine insufficiency in normal man. *Br J Nutr* 1987;58:207–14.
101. Rizzo C, Ribes A, Pastore A, Dionisi-Vici C, Greco M, Rizzoni G, et al. Pyroglutamic aciduria and nephropathic cystinosis. *J Inher Metab Dis* 1999;22:224–6.
102. Pitt JJ, Hauser S. Transient 5-oxoprolinuria and high anion gap metabolic acidosis: clinical and biochemical findings in eleven subjects. *Clin Chem* 1998;44:1497–503.
103. Abeling NG, van Gennip AH, Barth PG, van Cruchten A, Westra M, Wijburg FA. Aromatic L-amino acid decarboxylase deficiency: a new case with a mild clinical presentation and unexpected laboratory findings. *J Inher Metab Dis* 1998;21:240–2.
104. Bindel TH, Fennessey PV, Miles BS, Goodman SI. 4-Hydroxycyclohexane-1-carboxylic acid: an unusual compound isolated from the urine of children with suspected disorders of metabolism. *Clin Chim Acta* 1976;66:209–17.
105. Jone CM, Wu AH. An unusual case of toluene-induced metabolic acidosis. *Clin Chem* 1988;34:2596–9.
106. Liebich HM, Pickert A, Tetschner B. Gas chromatographic and gas chromatographic-mass spectrometric analysis of organic acids in plasma of patients with chronic renal failure. *J Chromatogr* 1984;289:259–66.